

## MORE THAN BONES: ASIDE JOBS OF VITAMIN D

### MAS QUE HUESOS: TRABAJOS APARTE O ADICIONALES DE LA VITAMINA D

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#### Abstract

Vitamin D contributes to maintaining calcium levels in the bloodstream and maintains a good health of our bones. This vitamin activates a cascade of intracellular responses mediated by its nuclear receptor, vitamin D receptor. Interestingly, vitamin D receptor is expressed in almost all cells in our body, suggesting vitamin D could have additional roles. In this article, information about vitamin D and tuberculosis, obesity, diabetes, and cancer are presented.

**Keywords:** vitamin D, receptor, intracellular, bones.

#### Resumen

La vitamina D contribuye a mantener los niveles de calcio en el torrente sanguíneo y mantiene una buena salud de nuestros huesos. Esta vitamina activa una cascada de respuestas intracelulares mediadas por su receptor nuclear, el receptor de vitamina D. Curiosamente, el receptor de vitamina D se expresa en casi todas las células de nuestro cuerpo, lo que sugiere que la vitamina D podría tener funciones adicionales. En este artículo se presenta información sobre la vitamina D y la tuberculosis, la obesidad, la diabetes y el cáncer.

**Palabras clave:** vitamina D, receptor, intracelular, huesos.



## INTRODUCTION

Vitamin D was discovered in 1967 when experiments were conducted to identify the responsible molecule present in cod liver oil that could cure rickets (McCollum, 1967). Since then, and with the chemical synthesis of its metabolites (Narwid et al., 1974a; Narwid et al., 1974b; Partridge et al., 1974), the study of vitamin D was oriented towards its physiological roles in calcium and phosphate homeostasis in relation to bone density (Stern et al., 1975).

Currently, it is well-accepted that vitamin D can be initially acquired from the diet (Hribar et al., 2022) or synthesized from cholesterol or ergosterol in the skin upon solar UV-B light exposure (Khanna et al., 2022). This vitamin D, or cholecalciferol or vitamin D<sub>3</sub>, is then transferred to the liver where it is converted to calcifediol or 25-hydroxycholecalciferol by hydroxylation of vitamin D<sub>3</sub> by the enzyme vitamin D 25-hydroxylase in the endoplasmic reticulum (Sulimovici et al., 1979). Afterwards, calcifediol is converted to calcitriol or 1,25-dihydroxyvitamin D in the kidney by the cytochrome P450 enzyme 1-alpha-hydroxylase (Miller & Portale, 2000). While the function of hepatic 25-hydroxylase is mainly unregulated, renal 1-alpha-hydroxylase is tightly regulated by the parathormone (Aarskog et al., 1983; Kremer & Goltzman, 1982) controlling the availability of the active circulating hormone calcitriol.

In circulation, cholecalciferol, calcifediol and calcitriol are transported by the vitamin D binding protein (DBP) (Bouillon et al., 2020). In the absence of DBP, the levels of total circulating calcitriol mainly diminish (Zella et al., 2008). Interestingly, DBP gene polymorphisms have been associated with alterations in bone density due to its role in vitamin D bioavailability (Bhan, 2014).

Calcitriol regulates calcium intestinal absorption by increasing the calcium transporter in the apical

membrane and calcium ATP-dependent pump in the baso-lateral membrane of the enterocyte (Wasserman & Fullmer, 1989). In addition, calcitriol promotes bone resorption to maintain the 2.5 mM calcium concentration in blood (Brown et al., 2002, p. 2).

Calcitriol activates the transcription of target genes due to its intracellular receptor, vitamin D receptor (VDR) (Carlberg, 2022). Upon binding to calcitriol, VDR dimerizes with retinoid X receptors to activate or repress the transcription of genes containing vitamin D response elements (Abbas, 2017). Importantly, VDR has been detected in almost all types of cells (Carlberg, 2022), opening the question of additional roles of vitamin D beyond bone health. Considering the different cell targets of vitamin D, in this mini-review, novel physiological impacts of vitamin D are presented and discussed.

## VITAMIN D AND TUBERCULOSIS

Tuberculosis (TB) is an airborne disease caused by the bacillus *Mycobacterium tuberculosis*. An estimated 10.6 million people fell ill with TB in 2021 (World Health Organization, 2022) and, without treatment, the death rate from TB disease is higher (about 50%) (Menzies et al., 2018). In 2021, there were an estimated 1.4 million deaths among HIV-negative people and 187 000 deaths among HIV-positive people, for a combined total of 1.6 million (World Health Organization, 2022). Some people develop no symptomatology nor spread the bacteria when they get contaged, this is called a latent TB infection (LTBI). It is estimated that 5-15% of the people with LTBI will progress to TB in their lifetime (Cao et al., 2022).

Vitamin D has been proposed to play a role in reducing the susceptibility to LTBI (Ganmaa et al., 2019), the spreading rate in persons that have contact with TB patients (Aibana et al., 2019) and the conversion in tuberculin skin test (Ganmaa

et al., 2012), among others (Martineau et al., 2007). However, the results obtained now are not conclusive and more studies are necessary to acknowledge the real role of Vitamin D in the TB infection.

#### **VITAMIN D AND OBESITY**

Due to the molecular composition of vitamin D, the adipose tissue has been considered as the main tissue for its storage (Mutt et al., 2014). Earlier, it was reported that there was a significant inverse correlation between serum calcitriol and body mass index, where obese subjects had significantly lower calcitriol than non-obese volunteers (Parikh et al., 2004). Thus, the idea of vitamin D negatively regulating adipogenesis was postulated using animal models (Wood, 2008). However, calcitriol was shown to promote adipose differentiation of human subcutaneous preadipocytes (Nimitphong et al., 2012) and to promote adipocyte differentiation of human mesenchymal cells with enhanced lipid accumulation (Narvaez et al., 2013).

To highlight the complex roles of vitamin D in obesity, it has been demonstrated that null expression of 1-alpha-hydroxylase caused lower leptin levels and consequently increased the consumption of food in mice (Narvaez et al., 2009). Recently, experiments on subcutaneous adipose tissue have demonstrated that calcitriol bound VDR inhibits an inflammatory response and adipokine expression in adipocytes (Nimitphong et al., 2021).

These data support the suggestion that vitamin D status may regulate obesity and obesity-related inflammation. Further clinical prospective studies are still required to better understand how vitamin D metabolism affects obesity.

#### **VITAMIN D AND DIABETES**

Initially it was proposed that calcitriol could regulate the glucose sensor machinery and stimulate insulin

secretion in pancreatic  $\beta$  cells *in vitro* (Bouckenooghe et al., 2003) and in a rat model (Kadowaki & Norman, 1985). Moreover, a meta-analysis study reported that vitamin D deficiency could increase the risk for type 2 diabetes mellitus (Pittas et al., 2007). However, one year supplementation of vitamin D had no effect on insulin levels of prediabetic patients in a randomized placebo controlled clinical trial (Davidson et al., 2013). Also, supplementation with vitamin D3 failed to reduce the risk of diabetes of volunteers with high risk of type 2 diabetes (Pittas et al., 2019). These controversial studies point to the need for more research on the connection between vitamin D and diabetes.

#### **VITAMIN D AND CANCER**

The interest in the role of vitamin D in carcinogenesis began when studies in leukemia cells showed that calcitriol could inhibit the promotion of tumor formation of the well-known carcinogenic agent 12-O-tetradecanoylphorbol-13-acetate (Kuroki & Suda, 1983). Since then, several studies have highlighted the role of vitamin D in cancer. In particular, animal models to study this relationship have strongly suggested that vitamin D and/or calcitriol can prevent or cure several types of cancer (VanWeelden et al., 1998; W. Zheng et al., 2012; Y. Zheng et al., 2011). However, it has been difficult to clearly demonstrate this relationship in humans. Thus, recent meta-analyses have shown that low serum calcifediol is associated with reduced survival among female breast cancer patients (Li et al., 2021) and that vitamin D intake can reduce the incidence of pancreatic cancer (Liu et al., 2018). Meanwhile, this association in human subjects has not been observed in prostate cancer (Gilbert et al., 2011) nor basal cell carcinoma of the skin (Hunter et al., 1992).

*In vitro* and animal studies demonstrate an important role of vitamin D in cancer development and progression. However, other factors seem to be present in human metabolism that modify this

relationship. Therefore, more studies are required to study the role of vitamin D metabolism in human cancer.

### CONCLUSION

Different types of normal cells express VDR and 1-alpha-hydroxylase, which indicate that vitamin D metabolism may have a plethora of responses in our body. In addition to the well-established role of vitamin D on calcium metabolism and bone health, interesting novel evidence supports the idea that this vitamin could be beneficial in other tissues and pathologies. More studies in humans are still required to study the impact of vitamin D and its metabolites in human health.

### BIBLIOGRAPHIC REFERENCES

- Aarskog, D., Aksnes, L., & Markestad, T. (1983). Effect of parathyroid hormone on cAMP and 1,25-dihydroxyvitamin D formation and renal handling of phosphate in vitamin D-dependent rickets. *Pediatrics*, *71*(1), 59–63.
- Abbas, M. (2017). Physiological functions of Vitamin D in adipose tissue. *The Journal of Steroid Biochemistry and Molecular Biology*, *165*(Pt B), 369–381. <https://doi.org/10.1016/j.jsbmb.2016.08.004>
- Aibana, O., Huang, C., Aboud, S., Arnedo-Pena, A., Becerra, M. C., Bellido-Blasco, J., Bhosale, R., Calderon, R., Chiang, S., Contreras, C., Davaasambuu, G., Fawzi, W., Franke, M., Galea, J., Garcia-Ferrer, D., Gil-Fortuño, M., Gomila-Sard, B., Gupta, A., Gupte, N., & Murray, M. (2019). Vitamin D status and risk of incident tuberculosis disease: A nested case-control study, systematic review, and individual-participant data meta-analysis. *PLoS Medicine*, *16*(9), e1002907. <https://doi.org/10.1371/journal.pmed.1002907>
- Bhan, I. (2014). Vitamin D Binding Protein and Bone Health. *International Journal of Endocrinology*, *2014*, 1–5. <https://doi.org/10.1155/2014/561214>
- Bouckenooghe, T., Vandewalle, B., Lukowiak, B., Kerr-Conte, J., Beläich, S., Gmyr, V., Dubois, M., Riachy, R., & Pattou, F. (2003). Modulation of Specific Beta Cell Gene (Re)Expression during In Vitro Expansion of Human Pancreatic Islet Cells. *Cell Transplantation*, *12*(7), 799–807. <https://doi.org/10.3727/000000003108747271>
- Bouillon, R., Schuit, F., Antonio, L., & Rastinejad, F. (2020). Vitamin D Binding Protein: A Historic Overview. *Frontiers in Endocrinology*, *10*, 910. <https://doi.org/10.3389/fendo.2019.00910>
- Brown, A., Finch, J., & Slatopolsky, E. (2002). Differential effects of 19-nor-1,25-dihydroxyvitamin D<sub>2</sub> and 1,25-dihydroxyvitamin D<sub>3</sub> on intestinal calcium and phosphate transport. *Journal of Laboratory and Clinical Medicine*, *139*(5), 279–284. <https://doi.org/10.1067/mlc.2002.122819>
- Cao, Y., Wang, X., Liu, P., Su, Y., Yu, H., & Du, J. (2022). Vitamin D and the risk of latent tuberculosis infection: A systematic review and meta-analysis. *BMC Pulmonary Medicine*, *22*(1), 39. <https://doi.org/10.1186/s12890-022-01830-5>
- Carlberg, C. (2022). Vitamin D and Its Target Genes. *Nutrients*, *14*(7), 1354. <https://doi.org/10.3390/nu14071354>
- Davidson, M., Duran, P., Lee, M., & Friedman, T. (2013). High-Dose Vitamin D Supplementation in People with Prediabetes and Hypovitaminosis D. *Diabetes Care*, *36*(2), 260–266. <https://doi.org/10.2337/dc12-1204>
- Ganmaa, D., Giovannucci, E., Bloom, B., Fawzi, W., Burr, W., Batbaatar, D., Sumberzul, N., Holick,

- M., & Willett, W. (2012). Vitamin D, tuberculin skin test conversion, and latent tuberculosis in Mongolian school-age children: A randomized, double-blind, placebo-controlled feasibility trial. *The American Journal of Clinical Nutrition*, *96*(2), 391–396. <https://doi.org/10.3945/ajcn.112.034967>
- Ganmaa, D., Khudyakov, P., Buyanjargal, U., Jargalsaikhan, B., Baigal, D., Munkhjargal, O., Yansan, N., Bolormaa, S., Lkhagvasuren, E., Sempos, C. T., Bromage, S., Wu, Z., Ochirbat, B., Gunchin, B., & Martineau, A. (2019). Prevalence and Determinants of QuantiFERON-Diagnosed Tuberculosis Infection in 9810 Mongolian Schoolchildren. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*, *69*(5), 813–819. <https://doi.org/10.1093/cid/ciy975>
- Gilbert, R., Martin, R. M., Beynon, R., Harris, R., Savovic, J., Zuccolo, L., Bekkering, G., Fraser, W., Sterne, J., & Metcalfe, C. (2011). Associations of circulating and dietary vitamin D with prostate cancer risk: A systematic review and dose-response meta-analysis. *Cancer Causes & Control: CCC*, *22*(3), 319–340. <https://doi.org/10.1007/s10552-010-9706-3>
- Hribar, M., Žlavs, K., Pravst, I., & Žmitek, K. (2022). Validation of the food frequency questionnaire for the assessment of dietary vitamin D intake. *Frontiers in Nutrition*, *9*, 950874. <https://doi.org/10.3389/fnut.2022.950874>
- Hunter, D., Colditz, G., Stampfer, M., Rosner, B., Willett, W., & Speizer, F. (1992). Diet and risk of basal cell carcinoma of the skin in a prospective cohort of women. *Annals of Epidemiology*, *2*(3), 231–239. [https://doi.org/10.1016/1047-2797\(92\)90055-u](https://doi.org/10.1016/1047-2797(92)90055-u)
- Kadowaki, S., & Norman, A. (1985). Demonstration that the Vitamin D Metabolite 1,25(OH)<sub>2</sub>-Vitamin D<sub>3</sub> and Not 24R,25(OH)<sub>2</sub>-Vitamin D<sub>3</sub> Is Essential for Normal Insulin Secretion in the Perfused Rat Pancreas. *Diabetes*, *34*(4), 315–320. <https://doi.org/10.2337/diab.34.4.315>
- Khanna, T., Shraim, R., Zarkovic, M., van Weele, M., van Geffen, J., & Zgaga, L. (2022). Comprehensive Analysis of Seasonal and Geographical Variation in UVB Radiation Relevant for Vitamin D Production in Europe. *Nutrients*, *14*(23), 5189. <https://doi.org/10.3390/nu14235189>
- Kremer, R., & Goltzman, D. (1982). Parathyroid hormone stimulates mammalian renal 25-hydroxyvitamin D<sub>3</sub>-1 $\alpha$ -hydroxylase *in vitro*. *Endocrinology*, *110*(1), 294–296. <https://doi.org/10.1210/endo-110-1-294>
- Kuroki, T., & Suda, T. (1983). Similarity and dissimilarity between phorbol ester tumor promoters and 1 $\alpha$ ,25-dihydroxyvitamin D<sub>3</sub>, an active form of vitamin D<sub>3</sub>. *Princess Takamatsu Symposia*, *14*, 89–99.
- Li, C., Li, H., Zhong, H., & Li, X. (2021). Association of 25-hydroxyvitamin D level with survival outcomes in female breast cancer patients: A meta-analysis. *The Journal of Steroid Biochemistry and Molecular Biology*, *212*, 105947. <https://doi.org/10.1016/j.jsbmb.2021.105947>
- Liu, Y., Wang, X., Sun, X., Lu, S., & Liu, S. (2018). Vitamin intake and pancreatic cancer risk reduction: A meta-analysis of observational studies. *Medicine*, *97*(13), e0114. <https://doi.org/10.1097/MD.00000000000010114>
- Martineau, A., Wilkinson, R., Wilkinson, K., Newton, S., Kampmann, B., Hall, B., Packe, G., Davidson, R., Eldridge, S., Maunsell, Z., Rainbow, S.,

- Berry, J., & Griffiths, C. (2007). A single dose of vitamin D enhances immunity to mycobacteria. *American Journal of Respiratory and Critical Care Medicine*, 176(2), 208–213. <https://doi.org/10.1164/rccm.200701-007OC>
- McCollum, E. V. (1967). The Paths to the Discovery of Vitamins A and D. *The Journal of Nutrition*, 91(suppl\_2), 11–16. [https://doi.org/10.1093/jn/91.2\\_Suppl.11](https://doi.org/10.1093/jn/91.2_Suppl.11)
- Menzies, N., Wolf, E., Connors, D., Bellerose, M., Sbarra, A., Cohen, T., Hill, A., Yaesoubi, R., Galer, K., White, P., Abubakar, I., & Salomon, J. (2018). Progression from latent infection to active disease in dynamic tuberculosis transmission models: A systematic review of the validity of modelling assumptions. *The Lancet Infectious Diseases*, 18(8), e228–e238. [https://doi.org/10.1016/S1473-3099\(18\)30134-8](https://doi.org/10.1016/S1473-3099(18)30134-8)
- Miller, W., & Portale, A. (2000). Vitamin D 1 $\alpha$ -Hydroxylase. *Trends in Endocrinology & Metabolism*, 11(8), 315–319. [https://doi.org/10.1016/S1043-2760\(00\)00287-3](https://doi.org/10.1016/S1043-2760(00)00287-3)
- Mutt, S., Hyppönen, E., Saarnio, J., Järvelin, M., & Herzig, K. (2014). Vitamin D and adipose tissue—more than storage. *Frontiers in Physiology*, 5, 228. <https://doi.org/10.3389/fphys.2014.00228>
- Narvaez, C., Matthews, D., Broun, E., Chan, M., & Welsh, J. (2009). Lean phenotype and resistance to diet-induced obesity in vitamin D receptor knockout mice correlates with induction of uncoupling protein-1 in white adipose tissue. *Endocrinology*, 150(2), 651–661. <https://doi.org/10.1210/en.2008-1118>
- Narvaez, C., Simmons, K., Brunton, J., Salinero, A., Chittur, S., & Welsh, J. (2013). Induction of STEAP4 correlates with 1,25-dihydroxyvitamin D<sub>3</sub> stimulation of adipogenesis in mesenchymal progenitor cells derived from human adipose tissue. *Journal of Cellular Physiology*, 228(10), 2024–2036. <https://doi.org/10.1002/jcp.24371>
- Narwid, T., Blount, J., Iacobelli, J., & Uskokovi, M. (1974a). Vitamin D<sub>3</sub> Metabolites III. Synthesis and X-ray analysis of 1,25-dihydroxycholesterol. *Helvetica Chimica Acta*, 57(3), 781–789. <https://doi.org/10.1002/hlca.19740570332>
- Narwid, T., Cooney, K., & Uskokovi, M. (1974b). Vitamin D<sub>3</sub> Metabolites II. Further Syntheses of 25-Hydroxycholesterol. *Helvetica Chimica Acta*, 57(3), 771–781. <https://doi.org/10.1002/hlca.19740570331>
- Nimitphong, H., Guo, W., Holick, M., Fried, S., & Lee, M. (2021). Vitamin D Inhibits Adipokine Production and Inflammatory Signaling Through the Vitamin D Receptor in Human Adipocytes. *Obesity (Silver Spring, Md.)*, 29(3), 562–568. <https://doi.org/10.1002/oby.23109>
- Nimitphong, H., Holick, M., Fried, S., & Lee, M. (2012). 25-hydroxyvitamin D<sub>3</sub> and 1,25-dihydroxyvitamin D<sub>3</sub> promote the differentiation of human subcutaneous preadipocytes. *PLoS One*, 7(12), e52171. <https://doi.org/10.1371/journal.pone.0052171>
- Parikh, S., Edelman, M., Uwaifo, G., Freedman, R., Semega-Janneh, M., Reynolds, J., & Yanovski, J. (2004). The Relationship between Obesity and Serum 1,25-Dihydroxy Vitamin D Concentrations in Healthy Adults. *The Journal of Clinical Endocrinology & Metabolism*, 89(3), 1196–1199. <https://doi.org/10.1210/jc.2003-031398>
- Partridge, J., Faber, S., & Uskokovi, M. (1974). Vitamin D<sub>3</sub> Metabolites I. Synthesis of

- 25-hydroxycholesterol. *Helvetica Chimica Acta*, 57(3), 764–771. <https://doi.org/10.1002/hlca.19740570330>
- Pittas, A., Dawson-Hughes, B., Sheehan, P., Ware, J., Knowler, W., Aroda, V., Brodsky, I., Ceglia, L., Chadha, C., Chatterjee, R., Desouza, C., Dolor, R., Foreyt, J., Fuss, P., Ghazi, A., Hsia, D., Johnson, K., Kashyap, S., Kim, S., Staten, M. (2019). Vitamin D Supplementation and Prevention of Type 2 Diabetes. *New England Journal of Medicine*, 381(6), 520–530. <https://doi.org/10.1056/NEJMoa1900906>
- Pittas, A., Lau, J., Hu, F., & Dawson-Hughes, B. (2007). The Role of Vitamin D and Calcium in Type 2 Diabetes. A Systematic Review and Meta-Analysis. *The Journal of Clinical Endocrinology & Metabolism*, 92(6), 2017–2029. <https://doi.org/10.1210/jc.2007-0298>
- Stern, P., Trummel, C., Schnoes, H., & Deluca, H. (1975). Bone resorbing activity of vitamin D metabolites and congeners in vitro: Influence of hydroxyl substituents in the A ring. *Endocrinology*, 97(6), 1552–1558. <https://doi.org/10.1210/endo-97-6-1552>
- Sulimovici, S., Roginsky, M., Duffy, J., & Pfeifer, R. (1979). Calciferol 25-hydroxylase activity in smooth and rough endoplasmic reticulum of rat liver. *Archives of Biochemistry and Biophysics*, 195(1), 45–52. [https://doi.org/10.1016/0003-9861\(79\)90325-4](https://doi.org/10.1016/0003-9861(79)90325-4)
- VanWeelden, K., Flanagan, L., Binderup, L., Tenniswood, M., & Welsh, J. (1998). Apoptotic regression of MCF-7 xenografts in nude mice treated with the vitamin D<sub>3</sub> analog, EB1089. *Endocrinology*, 139(4), 2102–2110. <https://doi.org/10.1210/endo.139.4.5892>
- Wasserman, R., & Fullmer, C. (1989). On the molecular mechanism of intestinal calcium transport. *Advances in Experimental Medicine and Biology*, 249, 45–65. [https://doi.org/10.1007/978-1-4684-9111-1\\_5](https://doi.org/10.1007/978-1-4684-9111-1_5)
- Wood, R. (2008). Vitamin D and adipogenesis: New molecular insights. *Nutrition Reviews*, 66(1), 40–46. <https://doi.org/10.1111/j.1753-4887.2007.00004.x>
- World Health Organization. (2022). *Global tuberculosis report 2022*.
- Zella, L., Shevde, N., Hollis, B., Cooke, N., & Pike, J. (2008). Vitamin D-Binding Protein Influences Total Circulating Levels of 1,25-Dihydroxyvitamin D<sub>3</sub> but Does Not Directly Modulate the Bioactive Levels of the Hormone in Vivo. *Endocrinology*, 149(7), 3656–3667. <https://doi.org/10.1210/en.2008-0042>
- Zheng, W., Wong, K., Zhang, Z., Dougherty, U., Mustafi, R., Kong, J., Deb, D., Zheng, H., Bissonnette, M., & Li, Y. (2012). Inactivation of the vitamin D receptor in APC(min/+) mice reveals a critical role for the vitamin D receptor in intestinal tumor growth. *International Journal of Cancer*, 130(1), 10–19. <https://doi.org/10.1002/ijc.25992>
- Zheng, Y., Zhou, H., Ooi, L., Snir, A., Dunstan, C., & Seibel, M. (2011). Vitamin D deficiency promotes prostate cancer growth in bone. *The Prostate*, 71(9), 1012–1021. <https://doi.org/10.1002/pros.21316>